

Reovirus-Specific Messenger Ribonucleoprotein Particles from Hela Cells

Jürgen Kreft

Biologisches Institut der Universität Stuttgart

Z. Naturforsch. **35 c**, 1046–1051 (1980); received May 14/August 13, 1980

Hela Cells, Reovirus, Messenger Ribonucleoprotein Particles, mRNP-Proteins, Electron Microscopy

When reovirus-infected Hela cells are incubated at 43 °C virus-specific messenger RNA is released from the polysomes. It accumulates free in the cytoplasm as messenger ribonucleoprotein particles (mRNPs). These particles have a sedimentation rate of about 50S and a buoyant density in CsCl of 1.42 g/cm³. Reovirus mRNPs contain, besides all three size classes of reovirus messenger RNA, the same spectrum of proteins found in the polysomal mRNPs from uninfected cells, plus two additional proteins with molecular masses of 70000 d and 110000 d, respectively. Electron microscopic examination of the reovirus mRNP fraction reveals specific Y-shaped structures with a total mean length of 0.5 µm.

Introduction

The genome of the reoviruses consists of double-stranded RNA which is enclosed within the virus capsid as ten segments falling into three size classes, named L, M and S [1, 2]. In infected host cells each of these segments is transcribed into single-stranded messenger RNA molecules [3] by a virus-associated transcriptase [4]. These viral mRNAs are translated by the host cell polysomes and can be released from these either *in vitro* by treatment with ethylenediamine tetraacetic acid (EDTA) [5, 6] or *in vivo* by blocking the initiation of protein synthesis [7]. In both cases the viral mRNA is found in the form of messenger ribonucleoprotein particles (reo-mRNPs), the bulk of which sediments around 50S in sucrose gradients [5–7]. These particles contain all three size classes (l, m, s) of reovirus mRNA, their protein composition has repeatedly been analysed, but with divergent results. In one report viral capsomere proteins have been found as major constituents [5], others found no virus-coded proteins at all [7], in a third report the presence of the non-capsid protein $\sigma 2A$ has been demonstrated [6].

Materials and Methods

Hela cells (S3) were grown in suspension culture with Eagles minimum essential medium supple-

Abbreviations: EDTA, ethylenediamine tetraacetic acid; mRNA, messenger ribonucleic acid; mRNP, messenger ribonucleoprotein particle; SDS, sodium dodecyl sulfate; Tris, tris (hydroxymethyl) aminomethane.

Reprint requests to Dr. Jürgen Kreft, Institut für Genetik und Mikrobiologie, Röntgenring 11, D-8700 Würzburg.

0341-0382/80/1100-1046 \$ 01.00/0

mented with 5% calf serum to a cell density of 5×10^5 cells/ml. Reovirus type 3 was originally isolated and classified by Dr. Schwöbel (Tübingen). Virus propagation, titer assays by plaque formation on mouse L cells or by hemagglutination and virus purification by density gradient centrifugation in cesium chloride have previously been described [8–10].

For the *in vivo* dissociation of polysomes the cells were incubated for 20 min at 43 °C [17], then poured on crushed frozen saline, washed once and resuspended in cold buffer A (10 mM Tris/HCl pH 7.4, 3 mM MgCl₂, 7 mM mercaptoethanol, 10 mM KCl). After 10 min swelling the cells were gently homogenized in a Dounce homogenizer, Triton X-100 was added to a final concentration of 1% and the nuclei were sedimented by centrifugation ($700 \times g$, 4 min, 4 °C). The supernatant was centrifuged ($30\,000 \times g$, 20 min, 4 °C) to remove membranes and mitochondria. This postmitochondrial supernatant was centrifuged through sucrose gradients as indicated in the legends to the figures.

The proteins from messenger ribonucleoprotein particles obtained by pelleting the appropriate fractions from sucrose gradients were solubilized by incubation of the mixture of these particles and contaminating ribosomes with pancreatic ribonuclease (2 µg/ml) for 20 min at 20 °C in buffer B (as buffer A, but with 500 mM KCl). The reaction mixture was layered on a cushion of 15% sucrose in buffer A and centrifuged in a truncated tube (2.2 cm, rotor SW 50, 45 000 rpm, 2.25 h, 4 °C). The upper third (0.6 ml) of the tube content was used for the protein analysis [11].



Dieses Werk wurde im Jahr 2013 vom Verlag Zeitschrift für Naturforschung in Zusammenarbeit mit der Max-Planck-Gesellschaft zur Förderung der Wissenschaften e.V. digitalisiert und unter folgender Lizenz veröffentlicht: Creative Commons Namensnennung-Keine Bearbeitung 3.0 Deutschland Lizenz.

Zum 01.01.2015 ist eine Anpassung der Lizenzbedingungen (Entfall der Creative Commons Lizenzbedingung „Keine Bearbeitung“) beabsichtigt, um eine Nachnutzung auch im Rahmen zukünftiger wissenschaftlicher Nutzungsformen zu ermöglichen.

This work has been digitalized and published in 2013 by Verlag Zeitschrift für Naturforschung in cooperation with the Max Planck Society for the Advancement of Science under a Creative Commons Attribution-NoDerivs 3.0 Germany License.

On 01.01.2015 it is planned to change the License Conditions (the removal of the Creative Commons License condition “no derivative works”). This is to allow reuse in the area of future scientific usage.

For electron microscopic examination the messenger ribonucleoprotein particles were suspended in buffer A, mixed with one volume of a cytochrome C solution (60 µg/ml in 0.3 M ammonium-acetate pH 7.1), a drop of 50 µl was pipetted on a paraffin sheet. After 5 min a parlodion/carbon coated grid (400 mesh) was brought in contact with the surface of the drop, then stained for 30 sec with uranylacetate (2×10^{-5} M in ethanol), washed once, dried and examined without any further treatment in a Zeiss EM 10 electron microscope at 60 kV.

The proteins from mRNPs were lyophilized, dissolved in sample buffer (10 mM Tris/HCl pH 7.4, 24 mM mercaptoethanol, 4% SDS, 10% glycerol), heated for 1 min at 90 °C and separated on discontinuous acrylamide slab gels [15]. The stacking gel contained 5% acrylamide in 125 mM Tris/HCl pH 7.8, the separation gel 10% acrylamide in 360 mM Tris/HCl pH 8.8, the electrode buffer 50 mM Tris/HCl pH 8.6, 385 mM glycine; the gels and the electrode buffer contained 0.2% SDS, electrophoresis was carried out at 75 mA. The gels were stained with coomassie blue and scanned at 580 nm. The molecular masses of the proteins were calculated by comparison with marker proteins [16].

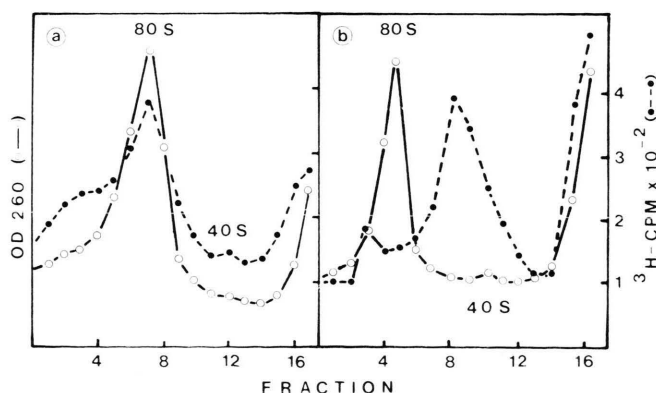
Further experimental details are given in the legends to the figures.

Results and Discussion

As found earlier in our laboratory arrest of protein synthesis and breakdown of polysomes can be induced in Hela cells by incubation at 43 °C for 20 min. After this treatment the bulk of cellular mRNA, rapidly labeled in the presence of 0.05 µg/ml actinomycin D, sediments in sucrose gradients

near the monoribosomal peak and represents polysomal messenger ribonucleoprotein particles (ps-mRNPs) [8, 17] (Fig. 1 a). The dissociation is reversible by lowering the temperature again to 37 °C and apparently does not lead to a damage of the ribosomes or the messenger ribonucleoprotein. At a concentration of 0.2 µg/ml actinomycin D the synthesis of cellular mRNA is almost completely blocked and the polysomes dissociate. Reovirus mRNA, however, continues to be synthesized at this concentration of actinomycin D and therefore can be selectively labeled [18]. The polysomes in reovirus-infected Hela cells also remain stable at this concentration of the drug, but can again be dissociated by incubation of the cells at 43 °C. Reovirus mRNA derived from the polysomes can be isolated from the postmitochondrial supernatant as a fraction sedimenting with 50S (Fig. 1 b). In non-infected cells treated in the same way no such peak at this position can be found. Since the largest species of reovirus mRNA sediments with 25S, the 50S fraction most likely contains the viral mRNA in a complexed form, *e.g.* associated with protein and/or ribosomal subunits. To reveal the nature of these complexes they were fixed with 3% glutaraldehyde [19] and banded in performed CsCl density gradients. Fig. 2 shows that the fixed particles have a mean buoyant density in CsCl of 1.42 g/cm³, from which a protein content of 70% can be estimated [20]. Such a density is characteristic for mRNPs, whereas complexes of mRNPs with the small ribosomal subunit have a density of 1.47 g/cm³ [17]. It must be noted that unfixed particles did not show any change in their sedimentation rate when they were centrifuged through sucrose gradients with concentrations of KCl up to 250 mM (results not

Fig. 1. Effect of elevated incubation temperature. Hela cells were infected with 15 PFU/cell of reovirus 3. After 14 h at 37 °C actinomycin D was added to these cells and to uninfected cells (0.2 and 0.05 µg/ml, respectively). At this time the synthesis of reovirus mRNA reaches a maximum. The cells were then labeled with 74 KBq/ml [³H]uridine for two h. The postmitochondrial supernatant was centrifuged through 15–30% sucrose gradients in buffer A (rotor SW 27, 200000 rpm, 17 h, 4 °C). Fractions of the gradients were analyzed for absorbance at 260 nm and for radioactivity precipitable with cold 5% trichloro acetic acid a) Uninfected cells; b) reovirus-infected cells.



shown). This is some evidence for the assumption that the 50S particles represent genuine complexes rather than artificial ones.

Analysis of the RNA and protein composition

Separation of purified RNA from the 50S fraction described above by gel electrophoresis shows that mainly RNA species with molecular masses of 0.4×10^6 , 0.7×10^6 and 1.3×10^6 d are present (Fig. 3). This corresponds well to the values given for the 14S (s), 18S (m) and 25S (l) species of reovirus mRNA [3]. The minor peaks migrating faster than the 0.4×10^6 d species must represent degradation products of reovirus mRNA since cellular RNA synthesis is almost completely blocked under the conditions which were used. This result clearly shows that the rapidly labeled 50S fraction, only found in reovirus-infected cells, consists of reovirus-specific mRNPs (reo-mRNPs).

Sucrose gradient fractions containing reo-mRNPs or ps-mRNPs are inevitably contaminated by ribosomes and ribosomal subunits. Unfortunately, since reovirus mRNA lacks a poly(A)-sequence [21], reo-mRNPs cannot be separated from the ribosomes by affinity chromatography, *e. g.* on poly(U)-sepharose.

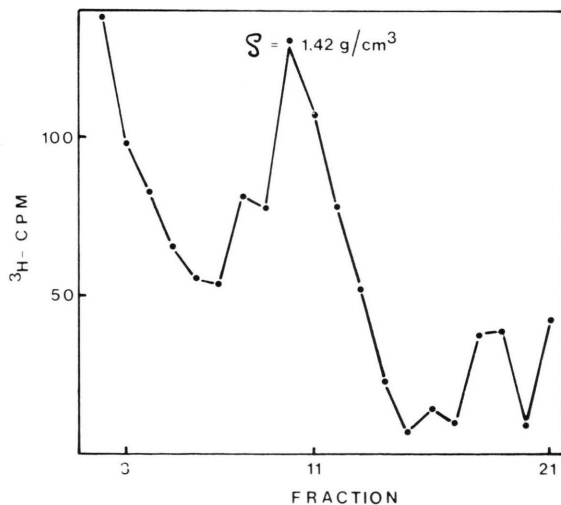


Fig. 2. Buoyant density of reo-mRNPs in CsCl. The 50S fraction from reovirus-infected Hela cells was prepared as described in Fig. 1 and pelleted in the rotor 50 Ti (45000 rpm, 16 h, 4 °C). The particles were resuspended in buffer A, fixed for 1 min at 4 °C with neutralized glutaraldehyde at a final concentration of 3% [19] and layered on preformed CsCl density gradients in buffer A (density 1.25–1.55 g/cm³). Centrifugation was in the rotor SW 56 at 35000 rpm for 16 h at 4 °C.

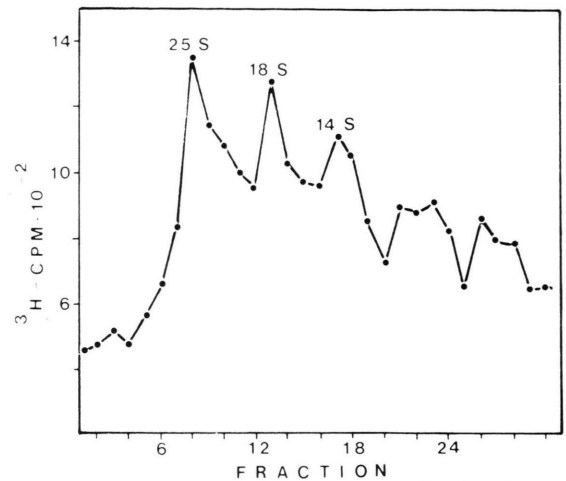


Fig. 3. RNA from reo-mRNPs. The 50S fraction from infected Hela cells was prepared as described in Fig. 1, the RNA was extracted with chloroform/phenol 1:1 [12] and analyzed by electrophoresis on polyacrylamide/agarose composite gels [13, 14]. The gels contained 2.2% acrylamide and 0.5% agarose in 90 mM Tris/HCl pH 8.3, 2.5 mM Na₂-EDTA, 0.15 M boric acid and 0.2% SDS. Unlabeled ribosomal RNA was used as a marker.

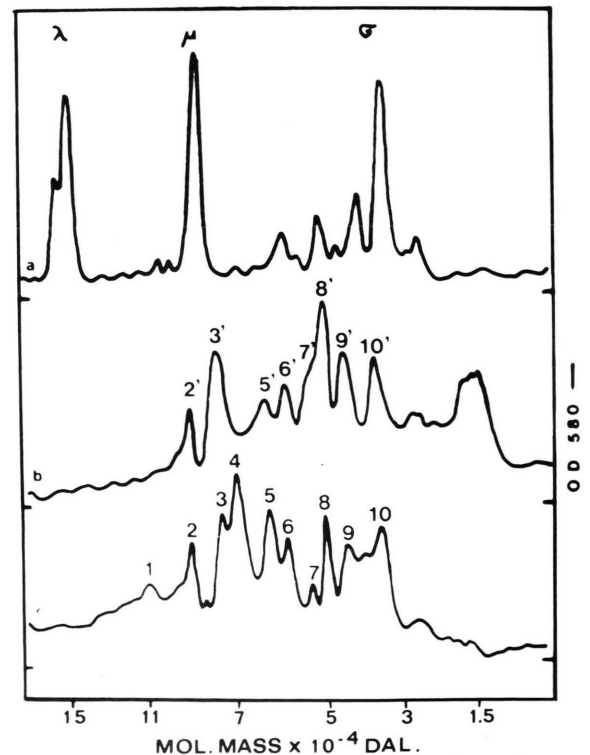


Fig. 4. Proteins of reovirus and messenger ribonucleoprotein particles. The analysis was performed on 10% polyacrylamide gels (see materials and methods), the stained gels were scanned at 580 nm. a) Purified reovirus; b) ps-mRNPs from uninfected cells; c) reo-mRNPs from reovirus-infected cells.

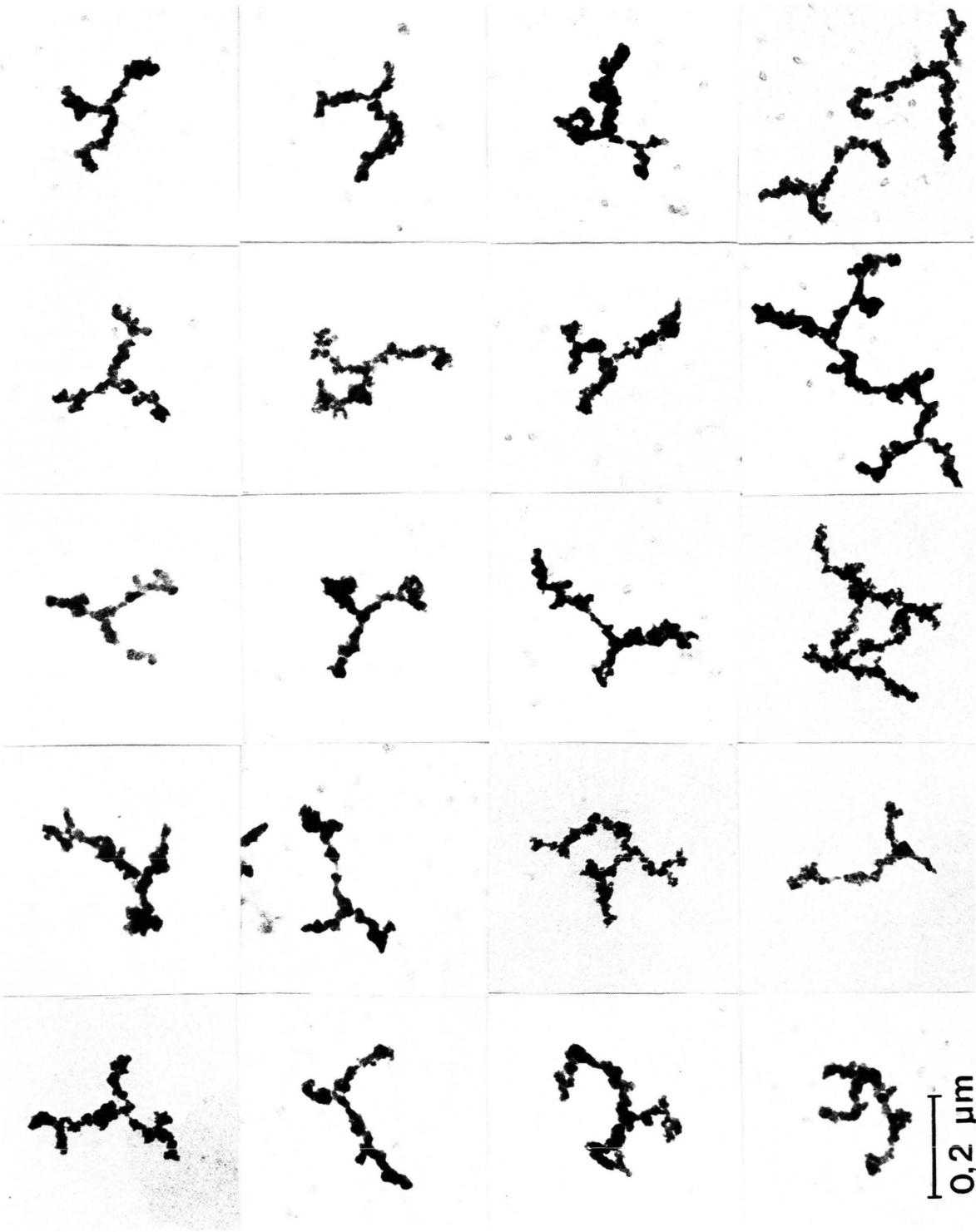


Fig. 5. Electron micrographs of reo-mRNPs from the 50S fraction of reovirus-infected cells. Three of the micrographs at the right show two particles each.

Table I. Molecular masses of the proteins from reovirus and from mRNPs ($\times 10^{-3} \pm 2000$ daltons). Numbers according to Fig. 4.

Reovirus	ps-mRNPs	reo-mRNPs
153		110 (1)
148	89 (2')	89 (2)
79	76 (3')	76 (3)
		70 (4)
	66 (5')	66 (5)
60	60 (6')	60 (6)
	54 (7')	54 (7)
48	48 (8')	48 (8)
39	43 (9')	43 (9)
34	36 (10')	36 (10)

It has been shown by others, however, that ribosomes are not degraded by low concentrations of pancreatic ribonuclease (2 $\mu\text{g}/\text{ml}$) while mRNPs are readily degraded under these conditions [22, 23]. Therefore mRNP proteins from non-infected and reovirus-infected Hela cells were solubilized by a limited RNase digestion of a mixture of mRNPs and ribosomes from which the latter were removed by centrifugation after the incubation with the enzyme. As was mentioned above the contamination of the reo-mRNP fraction with ps-mRNPs of cellular origin must be low since these have another sedimentation rate and since the cellular RNA and protein synthesis are strongly inhibited late in the infectious cycle [24] and at the concentration of actinomycin D which was used.

The proteins of purified virions, ps-mRNPs and reo-mRNPs were analyzed by gel electrophoresis. As can be seen from Fig. 4 and Table I reo-mRNPs contain a set of proteins of the same molecular masses as ps-mRNPs from non-infected Hela cells plus two additional proteins. One with a molecular mass of 110 000 d is present only in low amounts, the other with a molecular mass of 70 000 d is a main protein. It should be noted that adenovirus-infected KB cells also possess an additional 110 000 d protein in their ps-mRNPs [25]. None of the two additional proteins is a viral structural protein. In our experiments a wild strain of reovirus type 3 was used, the molecular weights of the viral proteins therefore differ somewhat from those of the more common Dearing strain. The presence of viral structural proteins in the 34 000–60 000 d region of the gels cannot be excluded by the method but no such proteins could be detected in the reo-mRNP fraction by immunoprecipitation with specific anti-

bodies. This is in agreement with the results of Christman *et al.* [7] and of Huismans and Joklik [6] but contradicts the results of Ward and Shatkin [5].

It has been reported that a protein of about 75 000 d binds specifically to the poly(A) sequences at the 3'-ends of most eukaryotic mRNAs [26]. Reovirus mRNA is devoid of poly(A) sequences at the 3'-end [23], the function of the 75 000 d protein in reo-mRNPs therefore remains to be elucidated. In histone mRNPs, which also contain no poly(A), a similar protein has recently been described [27].

Electron microscopy

The reo-mRNP fraction from sucrose gradients was further investigated by electron microscopy (Fig. 5). Numerous Y-shaped particles with a large, medium and a small branch and a total length of about 0.5 μm were found. No particles of this particular shape and size could be detected in uninfected Hela cells treated in the same way. Fig. 6 demonstrates that the large and the medium sized branches of the Y-shaped particles are rather homogeneous in length (an average of 0.22 μm and 0.16 μm , respectively), whereas the small branch ranges from 0.08–0.12 μm . The differences in the mean length of all three branches are statistically significant at the 99.5 percent level of probability. From these results it is concluded that the structures shown are specific and genuine ones rather than merely artefactual aggregates.

The extended lengths of the reo-mRNA species can be calculated to be 0.46 μm (s), 0.80 μm (m) and 1.49 μm (l), respectively, assuming that 0.86 Md are

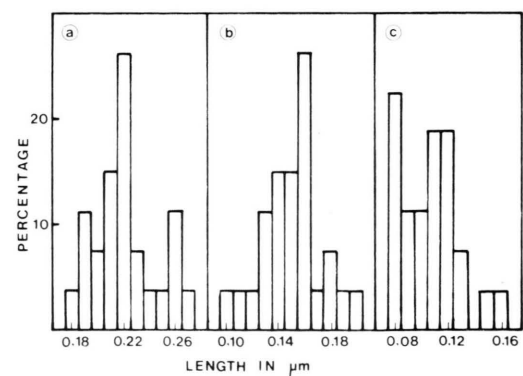


Fig. 6. Length distribution of the three branches of reo-mRNPs. The contour lengths were measured on large scale prints. a) Large branch; b) medium sized branch; c) small branch.

equivalent to 1 μm [28]. As only one class of reo-mRNPs was found by sedimentation analysis and electron microscopy, these particles most probably all contain the same assortment of reovirus mRNA molecules in a highly condensed or coiled form. Morphologically very similar but shorter (0.22 μm) and mostly linear structures have been depicted for globin mRNPs [29].

From all the results described above it becomes clear that in Hela cells infected with reovirus type 3 the viral mRNA is found in rather complex particles which have some properties in common with cellular mRNPs. Since these particles, the reo-mRNPs, can be isolated from the polysomal fraction of infected cells they are in fact engaged in the protein

synthesis. If they have in addition a function in the morphogenesis of the virions remains to be elucidated.

Acknowledgements

The author wishes to thank Prof. Dr. K. Köhler, who initiated this work, for his interest and support. He is grateful to him and to Dr. J.-P. Liautard for many discussions, to Drs. D. Hülser and H.-D. Royer for help with the electron microscope and to U. Tayebi and D. Tromm for their valuable technical assistance. This work was supported by a grant (to K. K.) from the Deutsche Forschungsgemeinschaft.

- [1] A. R. Bellamy, L. Shapiro, J. T. August, and W. K. Joklik, *J. Mol. Biol.* **29**, 1–18 (1967).
- [2] A. J. Shatkin, J. D. Sipe, and P. J. Loh, *J. Virol.* **2**, 986 (1968).
- [3] A. R. Bellamy and W. K. Joklik, *J. Mol. Biol.* **29**, 19–26 (1967).
- [4] J. Borsa and A. F. Graham, *Biochem. Biophys. Res. Commun.* **33**, 895–901 (1968).
- [5] R. L. Ward and A. J. Shatkin, *Arch. Biochem. Biophys.* **152**, 378–384 (1972).
- [6] H. Huismans and W. K. Joklik, *Virology* **70**, 411–424 (1976).
- [7] J. K. Christman, B. Reiss, D. Kyner, D. H. Levin, H. Klett, and G. Acs, *Biochim. Biophys. Acta* **294**, 153–164 (1973).
- [8] P. J. Gomatos and I. Tamm, *Virology* **17**, 455–461 (1962).
- [9] K. Köhler, *Z. Naturforsch.* **17 b**, 544–547 (1962).
- [10] S. J. Rhim and J. L. Melnick, *Virology* **15**, 80–81 (1961).
- [11] J.-P. Liautard, B. Setyono, E. Spindler, and K. Köhler, *Biochim. Biophys. Acta* **425**, 373–383 (1976).
- [12] S. Penman, *J. Mol. Biol.* **17**, 117–130 (1966).
- [13] C. W. Dingman and A. C. Peacock, *Biochemistry* **7**, 659–668 (1968).
- [14] A. C. Peacock and C. W. Dingman, *Biochemistry* **7**, 668–674 (1968).
- [15] U. K. Laemmli, *Nature* **227**, 680–685 (1970).
- [16] K. Weber and J. Osborn, *J. Biol. Chem.* **244**, 4406–4412 (1969).
- [17] G. Schochetman and R. P. Perry, *J. Mol. Biol.* **63**, 577–586 (1972).
- [18] A. J. Shatkin, *Biochem. Biophys. Res. Commun.* **19**, 506–510 (1965).
- [19] D. Baltimore and A. S. Huang, *Science* **162**, 572–574 (1968).
- [20] A. S. Spirin, *Europ. J. Biochem.* **10**, 20–35 (1969).
- [21] C. M. Stoltzfus, A. J. Shatkin, and A. K. Banerjee, *J. Biol. Chem.* **248**, 7993–7998 (1973).
- [22] J. R. Warner, P. Knopf, and A. Rich, *Proc. Nat. Acad. Sci. USA* **49**, 122–129 (1963).
- [23] S. Auerbach and T. Pederson, *Biochem. Biophys. Res. Commun.* **63**, 149–153 (1975).
- [24] R. L. Ward, A. K. Banerjee, A. La Fiandra, and A. J. Shatkin, *J. Virol.* **9**, 61–69 (1972).
- [25] U. Lindberg and B. Sundquist, *J. Mol. Biol.* **86**, 451–468 (1974).
- [26] G. Blobel, *Proc. Nat. Acad. Sci. USA* **70**, 924–928 (1973).
- [27] J.-P. Liautard and Ph. Jeanteur, *Nucl. Acids Res.* **7**, 135–150 (1979).
- [28] H. Delius, H. Westphal, and N. Axelrod, *J. Mol. Biol.* **74**, 677–687 (1973).
- [29] J. Dubochet, C. Morel, B. Lebleu, and M. Herzberg, *Europ. J. Biochem.* **36**, 465–472 (1973).